

## End User--- Proper BI Placement During VHP Decontamination Cycles

by Kurt McCauley

As is true with all sterilization processes, vaporized hydrogen peroxide (VHP) has clearly defined performance limits, many of which are unique to this process. VHP is primarily effective as a surface decontaminant and penetrates rather poorly as compared with other more traditional sterilization processes. Due to this limitation, proper biological indicator (BI) placement during the decontamination cycle is critical in order for the BI to function as designed.

The two main BI types used for monitoring the VHP cycles are a packaged disc and an unpackaged ribbon. A detailed description of these two BI types will be discussed as well as directions on their proper placement in the isolator. Additional information on their use is enclosed with each package of BI product.

### **The Packaged BI (HMV-091 series)**

The physical configuration of this BI is a stainless-steel disc inoculated with *Geobacillus stearothermophilus* spores packaged into a permeable Tyvek envelope. The spore placement is on the concave side of the disc, packaged with this side of the disc facing the printed side of the envelope.

When positioning the BIs in the isolator, the printed side should always face towards the vapor flow, and at no time should it be obstructed. Ideally the BI should be positioned such that the flow of vapor can pass by both sides of the envelope.

A hole has been provided in the envelope so the BI can be hung from a fixture on the wall, ceiling, or other structure. BI units can be hung individually or in multiples as demonstrated in Figure 1.



*Plastic spacers have been placed on the wire hanger between each BI to ensure proper vapor flow around each BI unit*



**Figure 1. Examples of proper BI configurations when using the hole in the envelope to hang the BI**

An adhesive tape can also be used to fix BIs into place. The best practice here is to place the tape on the peel flaps of the envelope and then secure it to the surface being monitored, as illustrated in Figure 2. Care must be taken not to cover the pocket of the envelope with the tape which would obstruct the vapor flow, as illustrated in Figure 3.

Note: Not all adhesive tapes are equal. Tape that absorbs or catalyzes hydrogen peroxide should not be used. Additionally, tape that leaves a residual behind once it is removed should be avoided.



**Figure 2. Examples of proper BI configurations when using adhesive tape to position the BI**



**Figure 3. Examples of IMPROPER BI configurations when using adhesive tape**

Another common practice, especially when using a large number of BIs, is to write identifying marks on each BI. These marks help identify the location in which the BI was placed as demonstrated in the first photograph in Figure 4. As is the case with the tape, placing marks on the pocket of the envelope should be avoided. Certain types of ink may also catalyze hydrogen peroxide and should be avoided completely.

Due to poor vapor flow, placing BIs into or underneath bottles (Figure 4) or other containers is not recommended.



**Figure 4. Examples of IMPROPER BI use: writing on the face of the package and placing BI into or underneath a container.**

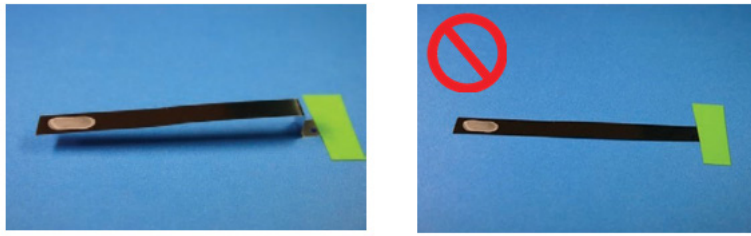
In large validations where hundreds of BIs are used in a single decontamination cycle, the BIs are often placed in the isolator the day prior to the exposure. Also prior to the exposure, a thorough cleaning or wipe down of the isolator may occur. When this happens, and especially if the isolator is sealed after the wipe down, the internal humidity may be high. It has been demonstrated that BIs stored under high humidity conditions may have an increased resistance to hydrogen peroxide. It is recommended that when both BI placement and isolator cleaning happen the day prior to exposure, the isolator is not sealed to prevent increased humidity levels.

### **The Unpackaged Ribbon BI (SBC-327 series)**

The physical configuration of this BI is a stainless-steel ribbon inoculated on one end with *Geobacillus stearothermophilus* spores. The inoculated area of the BI is clearly visible (a  $10^4$  spore load is still visible but not as obvious as a  $10^5$  or  $10^6$  load). The BIs are placed in a protective “cartridge belt” until the time of use.

Note: Sterile cartridge belts are available for transport of ribbons to the lab following exposure to sterilant.

The ribbon BI can also be positioned by hanging or taping in place. The inoculated area should always be positioned so that it faces outward and ideally so that the vapor can flow by both sides of the ribbon as demonstrated in Figure 5.



**Figure 5. Examples of proper (left photograph) and improper (right photograph) BI configurations**

An advantage of this BI is that the inoculated area of the ribbon can be placed into locations where the disc BI cannot physically fit, including restricted areas where the vapor penetration may be in question. Dead legs in an isolator are certainly areas of concern and should be eliminated or minimized. The ribbon can be useful in these cases in identifying the gas penetration down dead legs. The inoculated area can be placed into the area of concern, and if sufficient vapor flow occurs through this restriction, the spores on the BI will be inactivated. However, if the result after culturing the BI is “growth”, then it is fair to assume that inadequate vapor flow is occurring, and the area is a true dead leg. See Figure 6.

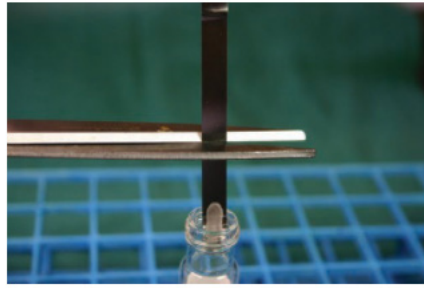


**Figure 6. Examples of ribbon BIs in dead leg (left) and hanging on an isolator fixture (right)**

### **Post exposure handling**

Post exposure BI handling is also important. In some cases, the BIs are cultured in the isolator, and in other cases, they are transported to a lab for culture. Tyvek is not known to absorb significant amounts of hydrogen peroxide, however when transporting the BIs to the lab, they should be placed into a porous bag (e.g., Tyvek) to allow for any out gassing. The positive control BIs can be transported to the lab at the same time, but they must never be placed into the same bag as the exposed units.

When culturing the ribbon BIs, only the inoculated end of the carrier should be cultured. This can be accomplished by cutting it free from the ribbon with sterile scissors as demonstrated in Figure 7. To culture the entire ribbon, especially if it has been held in place with tape, would invite false positive results.



**Figure 7. Example of culturing inoculated area of ribbon BI**

When a positive BI is obtained after a decontamination cycle, the results must be investigated. Was the result caused by a “hyper resistant” BI, poor BI positioning in the isolator, post exposure contamination, or perhaps the biological indicator is “indicating” there may truly be a problem in that particular location of the isolator? As was emphasized here, a few considerations in the pre-exposure set up and post-exposure handling should not be overlooked. As noted in a previous Spore News, the use of replicate BIs in problematic locations addresses the issue of the occasional “hyper resistant” BI.

Identifying the cause of a positive BI is not easy, but one thing is certain--If the BI is obstructed in any way, the vapor flow to the spores will be limited, and positive BI results can be expected.

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